

## **cis-trans-Isomerisation of the Proline-Peptide-Bond in a Cyclic Tetrapeptide Related to Chlamydocin\***

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Saturation transfer NMR experiments have been used to study the thermodynamic characteristics of the interconversion of two solute conformations of the chlamydocin derivative **2** in *DMSO-d*<sub>6</sub>.

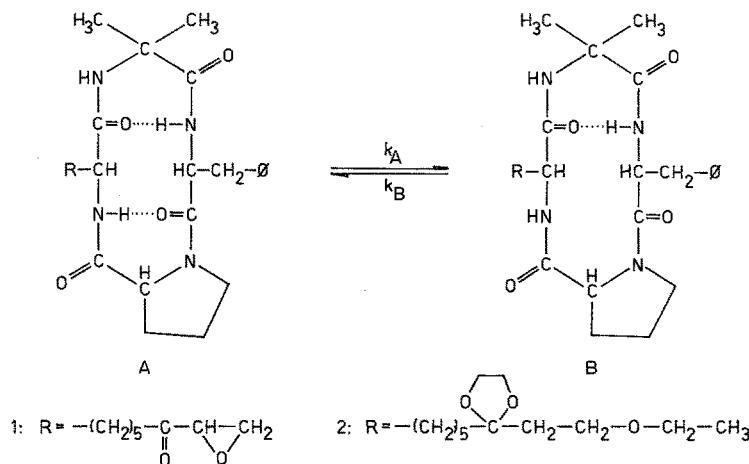
(Keywords: *Chlamydocin; Kinetics of conformational changes; Saturation transfer*)

*Cis-trans-Isomerisierung der Prolin-Peptidbindung in einem Derivat von Chlamydocin*

Durch Sättigungstransferexperimente wurde die Kinetik der Isomerisierung mittels NMR-Spektroskopie untersucht. Die thermodynamischen Größen der Reaktion wurden bestimmt.

The threedimensional structure of a peptide or protein can be related to biological functions of this molecule<sup>1</sup>. Therefore the conformational behaviour of small cyclic peptides was the aim of many investigations<sup>2-7</sup>. Chlamydocin (**1**), a cyclic tetrapeptide isolated from culture filtrates of *Diheterospora chlamydosporia*<sup>8</sup>, has been studied recently by NMR. It has been shown that chlamydocin and similar cyclic tetrapeptides exist in two conformations in *DMSO-d*<sub>6</sub><sup>5,6,9</sup>. We have studied the conformational behaviour of cyclo[A<sup>i</sup>bu-L-Phe-D-Pro-L-Ada], **2**, which is closely related to chlamydocin.

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The two conformers of **2** in  $DMSO-d_6$  have internal hydrogen bonds and interconvert slowly on the NMR time scale. One of these conformers is an all-*trans* conformation with two  $\gamma$ -turns from the amide proton of Phe to the carbonyl oxygen of Ada and from the amide proton of Ada to the carbonyl oxygen of Phe, respectively<sup>5,9</sup>. The second conformer has a *cis* Phe-Pro-amide bond<sup>5,6</sup>. The extreme downfield shift of the amide proton of Ada ( $\Delta\delta \sim 2.5$  ppm) in this conformer indicates an exposure of

Table 1. *Chemical shifts [ppm] and coupling constants [Hz] of the amide protons of the two conformers of **2** in  $DMSO-d_6$ ; peptide backbone angles  $\Phi_L$  [deg] derived from the coupling constants ( $\pm 10\%$ ); room temperature*

	A <sup>i</sup> bu		Phe		Ada	
	A	B	A	B	A	B
$\delta$	8.00	7.94	7.71	6.86	6.98	8.51
$^3J(\text{HNC}\alpha\text{H})$	—	—	11.2	7.6	11.2	8.8
	Ada		Phe			
	A	B	A	B		
$\Phi_L$	— 120	— 80	— 120	— 80		

Ada = ethylene ketal of 2-amino-10-ethoxy-8-oxo-decanoic acid (see formula).

$A^i$ bu = aminoisobutyric acid.

For definition of  $\Phi_L$  see Ref.<sup>10</sup>.

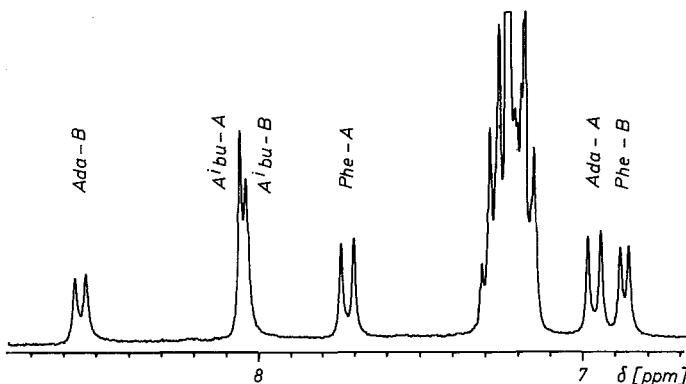


Fig. 1. Downfield part of the 250 MHz-<sup>1</sup>H-NMR-spectrum of **2** in  $DMSO-d_6$ . Amide resonances are marked according to conformers A and B.  $T = 305\text{ K}$

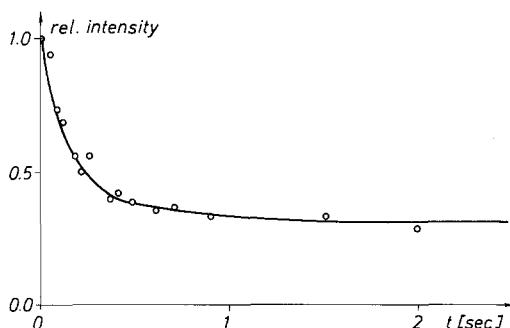


Fig. 2. Time dependence of the intensity of signal Ada-A upon instantaneous saturation of signal Ada-B at 325 K

this proton to the solvent and thus rupture of the  $\gamma$ -turn to the carbonyl oxygen of Phe during the conformational change. This is supported by the values of the coupling constants of the amide protons of Phe and Ada<sup>10,11</sup> (Tab. 1).

We have investigated the kinetics of the isomerisation by NMR double resonance techniques, first employed by *Hofmann* and *Forsén*<sup>12,13</sup>. These experiments have been done in the Pulse-FT-mode with the NH-protons of both isomers (Fig. 1). Instantaneous saturation of the transitions of one NH-proton was transferred to the signal of the corresponding NH-proton in the second isomer via the chemical exchange process, decreasing its intensity. The saturation transfer was

monitored as a function of time (Fig. 2) and yielded the magnetic relaxation times  $T_{1A}$  and  $T_{1B}$  of the corresponding protons and the values of  $k_A$  and  $k_B$  of the chemical exchange. Measurements of the rate constants  $k_A$  and  $k_B$  at various temperatures yielded the thermodynamic parameters of the isomerisation process (Tab. 2).

Table 2. *Kinetic and thermodynamic parameters of the isomerisation  $A \rightleftharpoons B$ .* We estimate the accuracy of the activation parameters to  $\pm 5\%$

$T$ [1]	$k_A$ [2]	$k_B$ [2]	$K$	$\Delta G$ [3]	$\Delta H$ [4]	$\Delta S$ [5]
295	1.62	1.67	0.97	75		-18
300	2.16	2.27	0.95	124		-18
305	3.08	3.48	0.89	310	-5.2	-18
315	5.24	6.13	0.85	411		-18

$T$ [1]	$\Delta G_A^*$ [4]	$\Delta G_B^*$ [4]	$\Delta H_A^*$ [4]	$\Delta H_B^*$ [4]	$\Delta S_A^*$ [5]	$\Delta S_B^*$ [5]
295	71.0	71.0			-86	-68
300	71.6	71.4			-86	-68
305	71.9	71.6	45.8	51.0	-86	-68
315	73.0	72.5			-86	-68

Units: [1]: K; [2]: s<sup>-1</sup>; [3]: J mol<sup>-1</sup>; [4]: kJ mol<sup>-1</sup>; [5]: J K<sup>-1</sup> mol<sup>-1</sup>.

$\Delta G^*$  of our study corresponds very well with  $\Delta G^*$  from earlier investigations which revealed values of 65 to 80 kJ mol<sup>-1</sup> for the *cis-trans*-isomerisation of X-Pro-bonds<sup>11,14</sup>.  $\Delta S^*$  of the conformational change is negative, suggesting a highly ordered transition state. Probably a solvent molecule is bound to the NH-proton of Ada supporting the rupture of the  $\gamma$ -turn. This too will contribute to a negative  $\Delta S^*$  as well as a high degree of order of the solvent molecules around the activated complex. This indicates the important role of the solvent during conformational changes of cyclic tetrapeptides.

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